Frozen Section Instructions

Contact: Christina Rotondi
HistoCore@mail.amc.edu  518-262-7090

Size of specimens:
1. Tissue should be trimmed to specified region before bringing to lab.
2. Clean off all fat and hair.
3. Size to fit embedding molds.
4. If cross sections are needed they should not protrude above embedding mold.

Preparation of tissue:
1. Freeze fresh tissue immediately on dry ice in an embedding mold using OCT.
2. Frozen tissue can be stored in -80°C freezer and transport on dry ice only. DO NOT use crushed ice.

Acceptable fixative:
1. 4% Paraformaldehyde (PFA) at 4°C -> place tissue in fixative immediately.
2. We recommend fixation of tissue samples for 24-48 hours; times will vary depending on size.
3. Incubate in 0.1M glycine (in dH2O) to remove an excess PFA for 1 hr at 4°C.
4. Incubate in 30% sucrose to cryoprotect tissue (in dH2O) overnight at 4°C. Samples will float at the beginning of the procedure and should sink when fully infiltrated.
5. Orient tissue samples in OCT after removing excess sucrose from surface of tissue.
6. Freeze sample on dry ice
7. Store at -80°C

Frozen sections:
1. Tissues will be cut at 5 – 10 microns, unless otherwise specified.
2. Two sections per slide unless otherwise specified.
3. Sections are placed on Superfrost® Plus slides.

Slide boxes:
1. Slide boxes need to be brought to the lab with tissue or will be provided at a charge.

NOTE:
- Please bring dry ice when picking up frozen tissue and prepared slides.
- A requisition form must be completed with all the required information.
- A brief consult with our staff will allow us to provide you with the best possible service.